# The ecological concept of costs of induced systemic resistance (ISR)

#### Martin Heil

Lehrstuhl Zoologie III, Biozentrum, Am Hubland D-97074 Würzburg, Germany (Phone: + 499318884378; Fax: +499318884352; E-mail: Martin\_Heil@hotmail.com); Centre d'Ecologie Fonctionelle et Evolutive (CEFE, CNRS) Route de Mende, 34293 Montpellier, Cédex 5, France

Accepted 18 October 2000

*Key words:* growth differentiation balance-hypothesis, fitness costs, induced defence, plant–pathogen interaction, SAR, systemic acquired resistance

#### **Abstract**

Plant defence is thought to provide benefits for the defended plants. Theoretical concepts must, therefore, explain why there is variation in defensive traits, which naively might be assumed to be present constitutively in fixed high amounts. Explanations are mainly based on the assumption of fitness costs. Investment in defence is thought to reduce the fitness of plants in enemy-free environments. Fitness costs often result from allocation costs, i.e. allocation of limited resources to defence, which then cannot be used for growth or other fitness-relevant processes. This theoretical concept can provide a useful tool for the interpretation of induced plant responses against pathogens, named induced systemic (or systemic acquired) resistance (ISR or SAR). Phenotypic plasticity, leading to induced responses, might have evolved mainly to reduce costs, since investment in defence is restricted to situations actually requiring defence. ISR can incur allocation costs and other, indirect costs, which ultimately may lead to fitness costs. Evolution of any defensive trait depends on both what a plant ideally 'should do' and what it actually 'is able to do'. Costs of defence constrain its expression. This might have important influences on the evolution of plant defensive traits, as well as on the exploitation of natural defences in agricultural crop protection.

Abbreviations: GDBH – growth differentiation balance hypothesis; JA – jasmonic acid; PR – pathogenesis related; SA – salicylic acid; SAR – systemic acquired resistance; ISR – induced systemic resistance.

## Introduction

Plants have evolved different strategies to defend themselves against herbivores and pathogens. Many of these strategies involve the induction of defensive traits, which therefore appear in high amounts or intensities only when plants suffer attack (Karban and Baldwin, 1997; Agrawal et al., 1999b; Tollrian and Harvell, 1999). Most studies on induced plant responses against herbivores have so far been conducted in an ecological context (Karban and Baldwin, 1997; Baldwin and Preston, 1999; Paul et al., 2000). In contrast, work on induced responses against pathogens has concentrated on (i) the specificity of interactions (Jackson and Taylor, 1996; Somssich and Hahlbrock, 1998),

(ii) the underlying signalling pathways (Ryals et al., 1994; Hunt et al., 1996; Hunt and Ryals, 1996; Schneider et al., 1996; van Loon, 1997), and (iii) how these findings might be used for agricultural purposes. Few studies have focused on ecological aspects of induced defences against pathogens (Heil, 1999; but see Hatcher, 1995). Some attempts have recently been made to integrate plant defence against herbivores and pathogens (Hatcher and Paul, 2000; Paul et al., 2000), but no unifying theory has been proposed for the ecological and evolutionary interpretation of both defensive strategies (Heil, 1999; 2000a).

In this review, I will argue that the ecological concept of costs, which is widely used in the context of antiherbivore defence, can also provide a useful framework to interpret findings on induced plant responses against pathogens. Studies on ISR that directly hint at relevant physiological costs will be reviewed along with evidence from studies in related fields such as, e.g., induced defence against herbivores, or direct defensive responses against pathogens. A much better understanding of the evolutionary and ecological constraints on induced responses is needed when these are to be used as effective methods in agricultural plant protection programmes.

## **Terminology**

The debate during the 'First International Symposium on Induced Resistance to Plant Diseases' held in Corfu in May 2000 revealed that much confusion exists as consequence of the use of different terms such as induced systemic resistance (ISR), systemic induced resistance (SIR), systemic acquired resistance (SAR), induced plant immunity, induced defence against pathogens, and so on. These terms are used synonymously by some authors, while others use them to describe different phenomena.

I follow the definitions given by Karban and Baldwin (1997) for induced defence against herbivores in naming all morphological or biochemical changes in plants that occur after some kind of attack or elicitor treatment 'induced responses', while those induced responses that affect the performance of the attackers negatively are named 'induced resistance'. The term 'induced defences' is used exclusively to denote those responses which have positive consequences for the fitness of the responding plants. While many studies have demonstrated that systemically induced responses of plants to pathogen attack successfully restrict further infections by the inducing as well as other pathogens (Ryals et al., 1994; Hammerschmidt and Kuć, 1995; Hoffland et al., 1996; Jackson and Taylor, 1996; Schneider et al., 1996; Anfoka and Buchenauer, 1997; and literature cited therein), no study has demonstrated so far that this response has a positive effect on plant fitness in a natural environment. Therefore, to date it has to be named resistance in the sense of Karban and Baldwin (1997). This term matches the original one, 'systemic acquired resistance', which had been chosen by Ross (1961).

The term 'acquired' seems to be the older one, and some arguments have been presented to favour this one over the term 'induced' (van Loon, 1997). Yet, all studies on responses of plants against insects use the

term 'induced', and an initial induction by a challenging infection or elicitor treatment is required for this specific form of systemic resistance against pathogens, too. This review will try to demonstrate that one framework can be used for the interpretation of plant defence against both pathogens and herbivores. A unified terminology helps to point to the similarities in both responses and to encourage further integration of concepts. I therefore name the whole set of changes by which plants respond to an initial infection, or elicitor treatment, in becoming systemically resistant against pathogen attack induced systemic resistance (ISR), and use this term synonymously with the other widespread term which is used to describe the same phenomenon, systemic acquired resistance (SAR).

# Evolutionary problems related to ISR and the concept of fitness costs

Induced systemic resistance in plants is a defensive mechanism that can be induced by a broad spectrum of pathogens, against many of which it is then effective (Hammerschmidt and Kuć, 1995). These main traits are already difficult to understand in an evolutionary or ecological context (Heil, 1999). What are the benefits of a systemically induced and unspecific resistance which relies on a 'challenging' infection? Why is ISR induced and not constitutive?

The concept of fitness costs has been used to explain why defensive traits that benefit plants are still subject to variation in natural populations, instead of being fixed genetically at a maximum level. This concept is also a useful tool to explain the occurrence of inducible defences, which are expressed only under certain circumstances. Stated simply, defence is assumed to incur costs to the plants, i.e., better defended plants are predicted to have a lower fitness as compared to less well defended plants, when both are compared under enemy-free conditions that prevent the defence from having any beneficial effects (Simms and Rausher, 1987; Simms and Fritz, 1990). Thus, defence should be established only when it is required. Although this explanation is widely used, surprisingly few studies have been conducted to measure fitness costs of any form of resistance, and only a restricted number of studies have demonstrated such costs (Bergelson and Purrington, 1996; but see Baldwin, 1998 and Agrawal et al., 1999a for the case of induced defence against herbivores, and Bergelson et al., 1996 for constitutive herbicide resistance). This has forced several authors

to propose alternative explanations as to why defence might be inducible (Agrawal and Karban, 1999).

Fitness costs may, but do not need to, be a consequence of allocation costs, which are addressed in most theoretical (e.g., Coley et al., 1985; Herms and Mattson, 1992) as well as empirical (for an overview, see e.g. Gershenzon, 1994) studies on costs of defences. Other consequences of defence, which might negatively affect plant fitness, can result from autotoxic properties of the defensive traits, or from negative influences on other organisms that affect plant fitness positively, i.e., mutualists. Moreover, even allocation costs translate to fitness costs only when the availability of resources is limited. Thus, they are not likely to appear under laboratory conditions, which are mostly characterised by optimised growing conditions and the absence of competition.

These problems may have prevented the allocation costs of ISR from becoming visible in most studies conducted so far, and only few studies have shown fitness costs of any form of pathogen resistance (but see Smedegaard-Petersen and Stolen, 1981; Smedegaard-Petersen and Tolstrup, 1985; Godard et al., 1999; Heil et al., 2000). However, many reported data, as well as theoretical considerations, are consistent with the assumption that ISR can incur relevant costs. This remains true for all three forms of costs to be discussed here, namely (i) allocation costs, (ii) costs resulting from autotoxicity of resistance traits, and (iii) costs resulting from negative influences on the plants' mutualists.

#### Allocation costs

All main steps of the signalling pathway, leading to ISR, require gene expression (Buell, 1999) and thus consume resources. This remains true for the initial steps characterised by the hypersensitive response (Baillieul et al., 1995; Kuć, 1995; Hunt and Ryals, 1996; Hammerschmidt and Nicholson, 1999), for the production of molecules such as salicylic acid (SA), which seem to be involved in signal transduction (Métraux et al., 1990; Mauch-Mani and Métraux, 1998; Hammerschmidt and Smith-Becker, 1999; Cameron, 2000), and for production of pathogenesis-related (PR) proteins (van Loon, 1997; van Loon and van Strien, 1999), phytoalexins (Kuć, 1995; Smith, 1996; Kuć, 1997; Hammerschmidt, 1999b; Hammerschmidt and Nicholson, van Loon and van Strien, 1999), and cell wall material (Nicholson and Hammerschmidt, 1992; Hammerschmidt and Nicholson, 1999), all of which form part of the plants' defensive responses. Consequently, Kuć (1995) has discussed whether stunted plants and lower productivity – that appear after application of elicitors of phytoalexin accumulation, or in plants which constitutively accumulate phytoalexins – may be caused by 'a marked diversion of energy and carbon precursors from vital processes'.

#### Autotoxicity

Some of the molecules involved in the signalling pathway (such as SA and the reactive oxygen species involved in the 'oxidative burst' which occurs at the beginning of the hypersensitive response, see Baker and Orlandi, 1995; Lamb and Dixon, 1997; Hammerschmidt and Nicholson, 1999) have already been demonstrated to have, or are likely to have, autotoxic effects (e.g., Rasmussen et al., 1991). Moreover, the hypersensitive response itself – which leads to controlled cell death at the site of infection – has at least some negative effects on the defending plant and thus presents a form of autotoxicity.

# Effects on mutualists

No study has so far demonstrated convincingly negative effects of ISR on any plant mutualist, but the low specificity of this defence and the high number of mutualistic interactions with microorganisms make it very likely that some of these might be negatively affected by ISR. Most plants have evolved mutualistic interactions with root-colonising fungi and bacteria, and many grasses have mutualistic, endosymbiotic fungi (Clay, 1990). It is already known that several interactions between mutualists infecting roots (i.e., mycorrhizal fungi and root nodulating bacteria) and the plants' induced resistance against pathogens do occur. Most of these interactions are characterised by an increase in the plants' defensive systems at the onset of the mutualistic infection (see, e.g., Dumas-Gaudot et al., 1996; Van Wees et al., 1997; Cordier et al., 1998; van Loon et al., 1998; Pieterse and Van Loon, 1999; Ruiz-Lozano et al., 1999). For example, infecting fungi must overcome plant defensive responses which occur during the establishment of mycorrhiza (Kapulnik et al., 1996), and establishment of mycorrhiza can be delayed in plants which constitutively express some (but not other) PR proteins (Vierheilig et al., 1995). Moreover,

chemical induction of pathogen resistance in alfalfa (*Medicago sativa*) and faba beans (*Vicia faba*) can lead to a reduced size and number of root nodules (Martínez-Abarca et al., 1998; Heil, 2000b). As a consequence, ISR might incur indirect costs by negatively affecting the plants' mutualistic interactions.

These studies are very preliminary, and most plant mutualists can be expected to be adapted to cope with the natural defensive responses of their host plant. Further studies are needed to determine whether findings derived from studies with artificial induction of ISR (Martínez-Abarca et al., 1998; Heil, 2000b) have any relevance for naturally elicited ISR. Yet, negative effects on mutualistic microorganisms might be a serious problem in agricultural systems which rely on chemical induction of ISR.

Moreover, some pathogenesis-related proteins have been reported to be induced as a consequence of attack by herbivorous insects (Mayer et al., 1996; Inbar et al., 1998; 1999), and these PR-proteins (e.g., chitinases) seem to have negative effects on herbivores (Broadway et al., 1998; Inbar et al., 1998; 1999). It has, therefore, already been hypothesised that chitinases could have negative effects on mutualistic insects, too (Heil et al., 1999).

#### Applying the concept of costs to ISR

In the following section, the concept of costs is used to formulate predictions on how different traits of ISR should be expressed. Several of these predictions have already been made by Herms and Mattson (1992) in the growth differentiation balance-hypothesis (GDBH), which is formulated in the context of constitutive defence against herbivores.

The GDBH is based on the main assumption that there exists competition between metabolic pathways relevant for growth and pathways involved in 'differentiation', with the latter term comprising all processes which are commonly called 'secondary metabolism': 'The GDBH of plant defence is premised upon a physiological trade-off between growth and differentiation processes. The trade-off between growth and defence exists because secondary metabolism and structural reinforcement are physiologically constrained in dividing and enlarging cells, and because they divert resources from the production of new leaf area' (Herms and Mattson, 1992). This trade-off clearly constrains the ability of plants to defend themselves against pathogen attack.

Besides the main assumption of metabolic competition, several clear predictions on when and how defensive traits should be expressed can be drawn from this concept: (i) plants cannot produce unlimited amounts of defensive compounds; (ii) young, developing plant parts are hard to defend by 'expensive' forms of defence which consume limited resources; (iii) negative interactions should become more obvious when plants suffer from a shortage of limiting resources such as, nutrients; (iv) plants should avoid redundant defences; (v) the production of defensive compounds can cause fitness costs.

#### Metabolic competition

Support for a 'metabolic competition' between primary metabolism and ISR comes from studies on potato and on parsley cell cultures, where Rubisco levels (potato) and the expression of histone-encoding genes (parsley) were strongly reduced after pathogen infection or elicitor treatment (Longemann et al., 1995; Somssich and Hahlbrock, 1998). 'The metabolic significance of gene repression concomitant with gene activation during pathogen defence is probably associated with the downregulation of all disposable cellular activities' (Somssich and Hahlbrock, 1998).

Additional studies have been conducted on differential gene induction and repression in response to methyljasmonate, an important elicitor of woundor herbivory-induced responses (for reviews, see Reinbothe et al., 1994; Creelman and Mullet, 1997). For example, Weidhase et al. (1987) reported the selective repression of several of those proteins which are present before jasmonate treatment. Reinbothe and co-workers have used barley to investigate responses of protein biosynthesis involved in production of, e.g., Rubisco and chlorophyll a/b binding proteins, to methyljasmonate treatment. Downregulation was found at the level of gene transcription (Reinbothe et al., 1994), in posttranscriptional transcript modifications (Reinbothe et al., 1993a), and in the stability of the transcripts (Reinbothe et al., 1993b). Finally, translation of mRNA encoding these proteins was also downregulated (Müller-Uri et al., 1988; Reinbothe et al., 1993b,c). Similar results have been presented in other studies (Wasternack et al., 1998).

Most of these results consider direct regulation processes rather than a simple metabolic 'competition' for limited resources. However, the question arises why biosynthesis involved in the production of vitally important proteins is downregulated at all. An answer can be derived from the assumption of resource limitations, which have to be coped with by controlled shifts in metabolic resource flows from primary metabolism to defence. Corresponding to this interpretation and to that given by Somssich and Hahlbrock (1998) (see above), Weidhase et al. (1987) and Reinbothe et al. (1994) proposed that the amino acids released by the proteolytic degradation of photosynthetic proteins are re-utilised for synthesis of defence proteins. These effects can finally lead to chlorophyll loss and Rubisco degradation (Weidhase et al., 1987; Reinbothe et al., 1994) and therewith form important costs of methyljasmonate-induced defence, which are likely to occur in a comparable way in ISR.

### Limited amounts of defence

Two main signalling pathways are involved in the induced plant responses against pathogens and herbivores. When plants are compromised in the total amounts of defence they can produce, negative interactions between these two responses are to be expected to occur, at least under limiting conditions. Several studies have already been conducted on reciprocal effects of induced responses against herbivores and pathogens. Some studies have provided hints on beneficial reciprocal influences. For example, herbivore-caused damage of tomato can increase resistance against the phytopathogen, Pseudomonas syringae (Stout et al., 1998b) and JA-inducible genes can be activated by pathogen attack (Schweizer et al., 1997). JA can induce phytoalexins and defence-related cell wall constituents (Nojiri et al., 1996; Creelman and Mullet, 1997; Wasternack and Parthier, 1997), and methyljasmonate can induce several PR-proteins (Xu et al., 1994). Treatment with both elicitors can protect tomato and potato against Phytophtora infestans (Cohen et al., 1993). Nicotine, which is induced as a response to herbivory (Baldwin, 1988a,b), can have negative effects on both herbivores and bacteria (Krischik et al., 1991). For overviews on reciprocally induced defences against pathogens and herbivores see Hatcher (1995) and Hatcher and Paul (2000).

However, most published data point to some kind of 'signalling conflicts' or 'trade-offs' (Bostock, 1999; Felton et al., 1999). The most thoroughly investigated system in this context is again tomato, *Lycopersicon esculentum*, for which both molecular and ecological studies have been conducted (for a review, see

Thaler, 1999). These studies have demonstrated that chemical induction of ISR decreases the plants' ability to express wound-inducible proteinase-inhibitors (Doares et al., 1995; Fidantsef et al., 1999). Correspondingly, treating leaves with a chemical elicitor of ISR increased their suitability for herbivorous caterpillars (Thaler et al., 1997; Stout et al., 1999). SA-treatment has been reported to inhibit wound- and JA-induced responses in the same plant (Stout et al., 1998c), and application of JA reduced the efficacy of chemical ISR elicitors (Thaler et al., 1997). Most probably, action of SA or related substances inhibits synthesis of JA (Penacortes et al., 1993).

There are striking theoretical problems in attempting to explain why an attack by one form of attacker has a predictive value for a reduced probability to face attacks by other organisms (Paul et al., 2000). The 'signalling conflicts' or 'trade-offs' between induced responses against pathogens and herbivores are, therefore, hard to explain when only the beneficial effects of the response are considered. However, evolution of any trait does not depend only on what a plant ideally 'should do', but also on factors constraining what it actually 'is able to do'. The costs of defence provide an easy explanation for the negative interactions between both defensive pathways. Plants might simply be compromised in the total amounts of defensive compounds which can be produced during a limited time interval.

## Defence and developmental stage

Few studies have investigated whether the ability of a plant, or part of a plant, to produce PR proteins or other ISR components depends on its age. The induction of PR-protein transcripts in tobacco leaf discs, floated on solutions of different sugars, depended strongly on the developmental stage of the leaves (Herbers et al., 1996), with older leaves being highly inducible, while the same concentration of sugars failed to elicit a clear response in leaves which had not fully unfolded. Herbers et al. (1996) used source-sink relationships for an interpretation and supposed that source leaves, which normally export sugars, suffer from a metabolic disturbance when facing external soluble sugars, while sink leaves are normally tailored to import external sugars, which therefore represent no metabolic disturbance. However, according to the hypothesis of Herms and Mattson (1992), young, still growing leaves may just be unable to produce PR-proteins simply because their whole metabolic apparatus is needed for the

biosynthesis of growth-relevant proteins. In contrast, other studies found that young, still growing leaves on intact plants are perfectly capable of producing PR proteins (van Loon, 1985). Further studies, focused directly on a possible age-dependency of production of ISR-relevant components, should be conducted to determine under which conditions ISR can be compromised in young, developing plant parts.

Reports on a further form of resistance against pathogens are in line with the hypothesis formulated above. Dependency of a resistance trait on leaf developmental stage which is comparable to that reported by Herbers et al. (1996) has been reported by Leisner et al. (1992; 1993) for resistance against long-distance movement of cauliflower mosaic virus in leaves of turnip and Arabidopsis. Correspondingly, the agerelated resistance of Arabidopsis occurs only in older, pre-senescent plants (Cameron, 2000). Similar temporal patterns of resistance in plants, or parts of plants, against pathogens increasing with developmental stage have been reported for maize (Moose and Sisco, 1994). The expression of these constitutive forms of defence may have evolved mainly according to ecological and evolutionary demands, i.e., better defence of plants which have reached the flowering stage. However, the described patterns are clearly consistent with the predictions of Herms and Mattson (1992) and Coley et al. (1985) that young plants, which, under natural conditions, are forced to grow very fast to compete successful with other plants, may be strongly compromised in their ability to defend themselves against pathogens or herbivores.

#### Resource availability

The ability of a plant to defend itself against enemies should depend on resource availability (Coley et al., 1985), and allocation costs of defence should have stronger effects on growth under limiting conditions. Most studies on this topic have been conduced in the context of induced defence against herbivores. For example, nitrogen supply can have strong influences of several inducible and constitutive defences of tomato (Stout et al., 1998a), and higher costs of induced nicotine production were found in plants growing on soils with lower nitrogen content (Baldwin et al., 1998) or under competitive conditions (Baldwin and Hamilton, 2000). The effect of chemical induction of ISR on growth and seed set was studied in wheat plants which were cultivated in pots under competitive

conditions and at different levels of nitrogen supply (Heil et al., 2000). Treating wheat plants with the chemical ISR elicitor BION® (benzo (1,2,3) thiadiazole-7-carbothioic acid S-methyl ester) under pathogen-free conditions resulted in a suppression of plant growth and a reduced yield, as compared to untreated plants. This suppression was most pronounced in plants suffering from a strong shortage of nitrogen (Heil et al., 2000). Comparable results have been obtained for wheat cultivated under agricultural field conditions by Stadnik and Buchenauer (1999), who reported that BION® treatment resulted in no positive effect on yield, most probably since the benefits of reduced infection rates were counterbalanced by the costs of ISR induction.

# Redundant defences

Little is known about whether a plants' own chemical defence against pathogens is reduced in cases where its function is performed by an alternative mechanism. Chitinase activities in so-called myrmecophytic plants, which are well defended against herbivores and pathogenic fungi by the action of mutalistic ants, were lower than in related plants which have no ant-defenders (Heil et al., 1999; submitted), supporting the prediction that redundant defences are reduced to avoid superfluous costs. Corresponding evidence comes from studies focused on the chemical anti-herbivore defence of myrmecophytic plants (Rehr et al., 1973; Seigler and Ebinger, 1987).

## Fitness costs

Studies on fitness costs of any resistance trait are scarce (but see Bergelson et al., 1996; Baldwin, 1998; Agrawal, 1999). However, as early as 1981, Smedegaard-Petersen and Stolen (1981) reported that a successful resistance response by barley against powdery mildew requires energy, and finally leads to a reduction in grain yield. It was later supposed that this energy requirement may be a reason for 'the limiting effect of disease resistance on yield' (Smedegaard-Petersen and Tolstrup, 1985). Negative effects on plant growth and flowering time of an overexpression of defence-related peroxidase in tobacco, and positive effects on the same fitness-relevant parameters of an underexpression of the same enzyme, have been reported by Lagrimini et al. (1997). Longemann et al. (1995) demonstrated reduced growth in parsley cell cultures that had been induced to express resistance

genes by infection or fungal elicitors: an effect that clearly would lead to reduced plant growth and fitness costs if it translates to the whole-plant level. Studies on chemically-induced ISR in wheat have shown a reduced seed set and, therewith, significant fitness costs of treatment with an ISR elicitor conducted under otherwise limiting conditions (Heil et al., 2000).

#### Conclusions and directions for future research

Most studies reported here have not been designed to test the hypothesis that resistance of plants against their enemies does cause costs. Nevertheless, they have revealed results which match predictions based on this hypothesis and thereby clearly corroborate it. Plants may evolve to reduce the costs of a distinct resistance trait (Bergelson and Purrington, 1996; Agrawal and Karban, 1999), and several factors that induce resistance may simultaneously have positive effects on the plants which counterbalance or even overweigh the costs of resistance. The latter seems to be the case when so-called plant-growth promoting bacteria induce pathogen resistance (van Loon et al., 1998).

The exact form of costs of resistance against pathogens, and whether they translate into evolutionary relevant fitness costs, does depend on both the type of defensive trait and the environmental conditions. These costs, which do not necessarily appear under all conditions, may nevertheless have strong influences on the evolution of defence, and thus on the plants' ability to express defence under certain circumstances.

This review strongly concentrated on the aspect of costs of ISR to demonstrate that allocation costs or other, indirect, forms of costs, that finally can translate to fitness costs of resistance, may have strong influences on the quality and quantity, and on the spatial and temporal patterns, of induced plant resistance against pathogens. This does not imply that other factors, which might influence the evolution of inducible defences (Agrawal and Karban, 1999), can be excluded from further debate. This is especially true for the argument that adaptation of pathogens or herbivores to a defensive trait is much more difficult when facing defences which are inducible and thus vary spatially and temporarily within populations and even within individual plants.

Even most arguments presented by Agrawal and Karban (1999) assume that plants cannot produce unlimited amounts of defence and thus are based

on the theory that defence does cause costs. The value of the argument that only few studies have so far demonstrated costs is biased severely by the general psychological problem that aspects of costs, such as reduced growth and seed set, or suppressed gene activity, are widely regarded as 'negative' results and thus are less likely to be published. This especially holds for applied studies. While much work has been done on the induction of gene expression after challenging infections or elicitor treatment, fewer studies have been reported on potential gene suppression or down-regulation (but see examples reviewed by Somssich and Hahlbrock, 1998). Correspondingly, 'stunted' phenotypes resulting from transformation experiments which have been conducted to produce defence-overexpressing plants would provide important information on the costs of that particular defence. They are, however, often regarded as some kind of failed experiments. These observations are seldom quantified and even more seldom published (personal communications from several authors). The downregulation of a gene is a result as good as its induction, and suppressive effects on plant growth are valuable results, too. Much more information on 'the other side of the coin', i.e., constraints that have influences on the expression of defence traits, would be important for both forms of research, basic research interested in the physiology, ecology and evolution of defence, and applied research devoted to the development of tools for agricultural plant protection.

#### Acknowledgements

I thank Rick Bostock, John Bailey, Leendert C. van Loon and the participants of the 1st postdoc meeting at the John Innes Centre (especially Chris Lamb, Catherine Feuillet and Giles Oldroyd) for many discussions which finally led to the formulation of this manuscript. A.A. Agrawal, K.E. Linsenmair, L.C. van Loon and Neil Oldham critically read the manuscript. Financial support by the DFG (TP C8, SFB 251 and grant He 3169/1-1) is gratefully acknowledged.

#### References

Agrawal AA (1999) Induced responses to herbivory in wild radish: effects on several herbivores and plant fitness. Ecology 80: 1713–1723

Agrawal AA and Karban R (1999) Why induced defenses may be favored over constitutive strategies in plants. In: Tollrian R and Harvell CD (eds) The ecology and evolution of inducible

- defenses (pp 45-61) Princeton University Press, Princeton, New Jersey, USA
- Agrawal AA, Strauss SY and Stout MJ (1999a) Costs of induced responses and tolerance to herbivory in male and female fitness components of wild radish. Evolution 53: 1093–1104
- Agrawal AA, Tuzun S and Bent E (1999b) *Inducible plant defenses against pathogens and herbivores: Biochemistry, ecology, and agriculture.* American Phytopathological Society Press, St. Paul, Minnesota, USA
- Anfoka G and Buchenauer H (1997) Systemic acquired resistance in tomato against *Phytophtora infestans* by pre-inoculation with tobacco necrosis virus. Phys Mol Plant Path 50: 85–101
- Baillieul F, Genetet I, Kopp M, Saindrenan P, Fritig B and Kauffmann S (1995) A new elicitor of the hypersensitive response in tobacco: A fungal glycoprotein elicits cell death, expression of defence genes, production of salicylic acid, and induction of systemic acquired resistance. Plant J 8: 551–560
- Baker CJ and Orlandi EW (1995) Active oxygen in plant pathogenesis. Annu Rev Phytopath 32: 299–321
- Baldwin IT (1988a) The alkaloidal responses of wild tobacco to real and simulated herbivory. Oecologia 77: 378–381
- Baldwin IT (1988b) Short-term damage-induced increases in tobacco alkaloids protect plants. Oecologia 75: 367–370
- Baldwin IT (1998) Jasmonate-induced responses are costly but benefit plants under attack in native populations. Proc Natl Acad Sci USA 95: 8113–8118
- Baldwin IT, Gorham D, Schmelz EA, Lewandowski CA and Lynds G (1998) Allocation of nitrogen to an inducible defense and seed production in *Nicotiana attenuata*. Oecologia 115: 541–552
- Baldwin IT and Hamilton WI (2000) Jasmonate-induced responses of *Nicotiana sylvestris* result in fitness costs due to impaired competitive ability for nitrogen. J Chem Ecol 26: 915–952
- Baldwin IT and Preston CA (1999) The eco-physiological complexity of plant responses to insect herbivores. Planta 208: 137–145
- Bergelson J and Purrington CB (1996) Surveying patterns in the cost of resistance in plants. Am Nat 148: 536–558
- Bergelson J, Purrington CB, Palm CJ and Lopez-Gutiérrez J-C (1996) Costs of resistance: A test using transgenic *Arabidopsis thaliana*. P Roy Soc Lond B Biol 163: 1659–1663
- Bostock RM (1999) Signal conflicts and synergies in induced resistance to multiple attackers. Phys Mol Plant Path 55: 99–109
- Broadway RM, Gongora C, Kain WC, Sanderson JP, Monroy JA, Bennett KC, Warner JB and Hoffmann MP (1998) Novel chitinolytic enzymes with biological activity against herbivorous insects. J Chem Ecol 24: 985–998
- Buell CR (1999) Genes involved in plant-pathogen interactions. In: Agrawal AA, Tuzun S, and Bent E (eds) Induced plant defenses against pathogens and herbivores: Biochemistry, Ecology, and Agriculture (pp 73–93) The American Phytopathological Society Press, St. Paul, Minnesota, USA
- Cameron RK (2000) Salicylic acid and its role in plant defense responses: what do we really know? Phys Mol Plant Path 56: 91–93
- Clay K (1990) Fungal endophytes of grasses. Annu Rev Ecol Syst 21: 275–297

- Cohen Y, Gisi U and Niderman T (1993) Local and systemic protection against *Phytophtora infestans* induced in potato and tomato plants by jasmonic acid and jasmonic methyl-ester. Phytopathology 83: 1054–1062
- Coley PD, Bryant JP and Chapin FS, III. (1985) Resource availability and plant antiherbivore defense. Science 230: 895–899
- Cordier C, Pozo MJ, Barea JM, Gianinazzi S and Gianinazzi-Pearson V (1998) Cell defense responses associated with localized and systemic resistance to *Phytophthora parasitica* induced in tomato by an arbuscular mycorrhizal fungus. Mol Plant-Microbe In 11: 1017–1028
- Creelman RA and Mullet JE (1997) Biosynthesis and action of jasmonates in plants. Annu Rev Plant Phys Plant Mol Biol 48: 355–381
- Doares SH, Narvaez-Vasquez J, Conconi A and Ryan CA (1995) Salicylic acid inhibits synthesis of proteinase-inhibitors in tomato leaves induced by systemin and jasmonic acid. Plant Phys 108: 1741–1746
- Dumas-Gaudot E, Slezack S, Dassi B, Pozo MJ, Gianinazzi-Pearson V and Gianinazzi S (1996) Plant hydrolytic enzymes (chitinases and beta-1,3-glucanases) in root reactions to pathogenic and symbiotic microorganisms. Plant and Soil 185: 211–221
- Felton GW, Korth KL, Bi JL, Wesley SV, Huhman DV, Mathews MC, Murphy JB, Lamb C and Dixon RA (1999) Inverse relationship between systemic resistance of plants to microorganisms and to insect herbivory. Current Biology 9: 317–320
- Fidantsef AL, Stout MJ, Thaler JS, Duffey SS and Bostock RM (1999) Signal interactions in pathogen and insect attack: expression of lipoxygenase, proteinase inhibitor II, and pathogenesis-related protein P4 in the tomato, *Lycopersicon esculentum*. Phys Mol Plant Path 54: 97–114
- Gershenzon J (1994) The cost of plant chemical defense against herbivores: a biochemical perspective. In: Bernays EA (ed) Insect-plant interactions (pp 105–173) CRC Press, Boca Raton
- Godard JF, Ziadi S, Monot C, Le Corre D and Silue D (1999) Benzothiadiazole (BTH) induces resistance in cauliflower (*Brassica oleracea* var *botrytis*) to downy mildew of crucifers caused by *Peronospora parasitica*. Crop Protection 18: 379–405
- Hammerschmidt R (1999a) Induced disease resistance: how do induced plants stop pathogens? Phys Mol Plant Path 55: 77–84
- Hammerschmidt R (1999b) Phytoalexins: what have we learned after 60 years? Annu Rev Phytopathol 37: 285–306
- Hammerschmidt R and Kuć J (1995) Induced resistance to disease in plants. Kluwer, Dordrecht, The Netherlands
- Hammerschmidt R and Nicholson RL (1999) A survey of defense responses to pathogens. In: Agrawal AA, Tuzun S, and Bent E (eds) Induced plant defenses against pathogens and herbivores: Biochemistry, Ecology, and Agriculture (pp 55–71) The American Phytopathological Society Press, St. Paul, Minnesota
- Hammerschmidt R and Smith-Becker JA (1999) The role of salicylic acid in disease resistance. In: Agrawal AA, Tuzun S, and Bent E (eds) Induced plant defenses against pathogens and herbivores: Biochemistry, Ecology, and Agriculture (pp 37–53)
   The American Phytopathological Society Press, St. Paul, Minnesota
- Hatcher PE (1995) Three-way interactions between plant pathogenic fungi, herbivorous insects and their host plants. Biol Reviews 70: 639–694

- Hatcher PE and Paul ND (2000) On integrating molecular and ecological studies of plant resistance: variety of mechanisms and breadth of antagonists. J Ecol 88: 702–706
- Heil M (1999) Systemic acquired resistance available information and open ecological questions. J Ecol 87: 341–346
- Heil M (2000a) Different strategies for studying ecological aspects of systemic acquired resistance (SAR). J Ecol 88: 707–708
- Heil M (2000b) On the costs of chemically induced resistance against pathogens. Page 83. First international symposium induced resistance to plant diseases, Corfu, Greece.
- Heil M, Fiala B, Boller T and Linsenmair KE (1999) Reduced chitinase activities in ant plants of the genus *Macaranga*. Naturwissenschaften 86: 146–149
- Heil M, Hilpert A, Kaiser W and Linsenmair KE (2000) Reduced growth and seed set following chemical induction of pathogen defence – does systemic acquired resistance (SAR) incur allocation costs? J Ecol 88: 645–654
- Heil M, Staehelin C and McKey D. Low chitinase activity in *Acacia* myrmecophytes and the costs of direct antifungal defence. Naturwissenschaften (in press)
- Herbers K, Meuwly P, Métraux J-P and Sonnewald U (1996) Salicylic acid-independent induction of pathogenesis-related protein transcripts by sugars is dependent on leaf developmental stage. FEBS Letters 397: 239–244
- Herms DA and Mattson WJ (1992) The dilemma of plants: to grow or to defend. Q Rev Biol 67: 283–335
- Hoffland E, Hakulinen J and van Pelt JA (1996) Comparison of systemic resistance induced by avirulent and nonpathogenic *Pseudomonas* species. Phytopathology 86: 757–762
- Hunt MD, Neuenschwander UH, Delaney TP, Weymann KB, Friedrich LB, Lawton KA, Steiner H-Y and Ryals JA (1996) Recent advances in systemic acquired resistance research a review. Gene 179: 89–95
- Hunt MD and Ryals JA (1996) Systemic acquired resistance signal transduction. Crit Rev Plant Sci 15: 583–606
- Inbar M, Doostdar H, Leibee GL and Mayer RT (1999) The role of plant rapidly induced responses in asymmetric interspecific interactions among insect herbivores. J Chem Ecol 25: 1961–1979
- Inbar M, Doostdar H, Sonoda RM, Leibee GL and Mayer RT (1998) Elicitors of plant defensive systems reduce insect densities and disease incidence. J Chem Ecol 24: 135–149
- Jackson AO and Taylor CB (1996) Plant-microbe interactions: life and death at the interface. Pl Cell 8: 1651–1668
- Kapulnik Y, Volpin H, Itzhaki H, Ganon D, Galili S, David R, Shaul O, Elad Y, Chet I and Okon Y (1996) Suppression of defence responses in mycorrhizal alfalfa and tobacco roots. New Phytol 133: 59–64
- Karban R and Baldwin IT (1997) Induced responses to herbivory. University of Chicago Press, Chicago and London
- Krischik VA, Goth RW and Barbosa P (1991) Generalized plant defense: effects on multiple species. Oecologia 85: 562–571
- Kuć J (1995) Phytoalexins, stress metabolism, and disease resistance in plants. Annu Rev Phytopathol 33: 275–297
- Kuć J (1997) Molecular aspects of plant responses to pathogens. Acta Physiologiae Plantarum 19: 551–559
- Lagrimini LM, Gingas V, Finger F, Rothstein S and Liu T-TY (1997) Characterization of antisense transformed plants

- deficient in the tobacco anionic peroxidase. Plant Phys 114: 1187–1196
- Lamb C and Dixon RA (1997) The oxidative burst in plant disease resistance. Annu Rev Plant Phys Plant Mol Biol 48: 251–275
- Leisner SM, Turgeon R and Howell SH (1992) Long distance movement of cauliflower mosaic virus in infected plants. Mol Plant-Microbe In 5: 41–47
- Leisner SM, Turgeon R and Howell SH (1993) Effects of host plant development and genetic determinants on the long distance movement of cauliflower mosaic virus in *Arabidopsis*. Pl Cell 5: 191–202
- Longemann E, Wu S-C, Schröder J, Schmelzer E, Somssich IE and Hahlbrock K (1995) Gene activation by UV light, fungal elicitor or fungal infection in *Petroselium crispum* is correlated with repression of cell-cycle-related genes. Plant J 8: 865–876
- Martínez-Abarca F, Herrera-Cervera JA, Bueno P, Sanjuan J, Bisseling T and Olivares J (1998) Involvement of salicylic acid in the establishment of the *Rhizobium meliloti*-alfalfa symbiosis. Mol Plant-Microbe In 11: 153–155
- Mauch-Mani B and Métraux J-P (1998) Salicylic acid and systemic acquired resistance to pathogen attack. Ann Bot 82: 535–540
- Mayer RT, McCollum TG, McDonald RE, Polston JE and Doostdar H (1996) *Bemisia* feeding induces pathogenesis-related proteins in tomato. In: Gerling D and Mayer RT (eds) *Bemisia* 1995: Taxonomy, Biology, Damage Control and Management (pp 179–188) Intercept Ltd., Andover, Hants, UK
- Métraux JP, Signer H, Ryals J, Ward E, Wyss-Benz M, Gaudin J, Raschdorf K, Schmid E, Blum W and Inverardi B (1990) Increase in salicylic acid at the onset of systemic acquired resistance. Science 250: 1004–1006
- Moose SP and Sisco PH (1994) *Glossy* 15 controls the epidermal juvenile-to adult phase transition in maize. Plant Cell 6: 1343–1355
- Müller-Uri F, Parthier B and Nover L (1988) Jasmonate-induced alteration of gene expression in barley leaf segments analyzed by *in-vivo* and *in-vitro* protein synthesis. Planta 176: 241–247
- Nicholson RL and Hammerschmidt R (1992) Phenolic compounds and their role in disease resistance. Annu Rev Phytopathol 30: 369–389
- Nojiri H, Sugimori M, Yamane H, Nishimura Y, Yamada A, Shibuya N, Kodama O, Murofushi N and Omori T (1996) Involvement of jasmonic acid in elicitor-induced phytoalexin production in suspension-cultured rice cells. Plant Phys 110: 387–392
- Paul ND, Hatcher PE and Taylor JE (2000) Coping with multiple enemies: an integration of molecuar and ecological perspectives. Tr Plant Sci 5: 220–225
- Penacortes H, Albrecht T, Prat S, Weiler EW and Willmitzer L (1993) Aspirin prevents wound-induced gene-expression in tomato leaves by blocking jasmonic acid biosynthesis. Planta 191: 123–128
- Pieterse CMJ and Van Loon LC (1999) Salicylic acid-independent plant defence pathways. Tr Plant Sci 4: 52–58
- Rasmussen JB, Hammerschmidt R and Zook MN (1991) Systemic induction of salicylic acid accumulation in cucumber after inoculation with *Pseudomonas syringae* pv *syringae*. Plant Phys 97: 1342–1347

- Rehr SS, Feeny PP and Janzen DH (1973) Chemical defence in Central American non-ant Acacias. J Anim Ecol 42: 405–416
- Reinbothe S, Mollenhauer B and Reinbothe C (1994) JIPs and RIPs: the regulation of plant gene expression by jasmonates in response to environmental cues and pathogens. Plant Cell 6: 1197–1209
- Reinbothe S, Reinbothe C, Heintzen C, Seidenbecher C and Parthier P (1993a) A methyl jasmonate-induced shift in the length of the 5' untranslated region impairs translation of the plastid rbcL transcript in barley. EMBO J 12: 1505–1512
- Reinbothe S, Reinbothe C and Parthier B (1993b) Methyl jasmonate-regulated translation of nuclear-encoded chloroplast proteins in barley (*Hordeum vulgare* L. cv. salome). J Biol Chem 268: 10606–10611
- Reinbothe S, Reinbothe C and Partier B (1993c) Methyl jasmonate represses translation initiation of a specific set of mRNAs in barley. Plant J 4: 459–467
- Ross AF (1961) Systemic acquired resistance induced by localized virus infection in plants. Virology 14: 340–358
- Ruiz-Lozano JM, Roussel H, Gianinazzi S and Gianinazzi-Pearson V (1999) Defense genes are differentially induced by a mycorrhizal fungus and *Rhizobium* sp. in wild-type and symbiosis-defective pea genotypes. Mol Plant-Microbe In 12: 976–984
- Ryals J, Uknes S and Ward E (1994) Systemic acquired resistance. Plant Phys 104: 1109–1112
- Schneider M, Schweizer P, Meuwly P and Métraux JP (1996) Systemic acquired resistance in plants. In: Jeon KW (ed) Int Rev Cytology (pp 303–340) Academic Press, San Diego, California, USA
- Schweizer P, Buchala A, Silverman P, Seskar M, Raskin I and Metraux JP (1997) Jasmonate-inducible genes are activated in rice by pathogen attack without a concomitant increase in endogenous jasmonic acid levels. Plant Phys 114: 79–88
- Seigler DS and Ebinger JE (1987) Cyanogenic glycosides in antacacias of Mexico and Central America. SW Nat 32: 499–503
- Simms EL and Fritz RS (1990) The ecology and evolution of host-plant resistance to insects. Tr Ecol Evol 5: 356–360
- Simms EL and Rausher MD (1987) Costs and benefits of plant resistance to herbivory. Am Nat 130: 570–581
- Smedegaard-Petersen V and Stolen O (1981) Effect of energy requiring defense reactions on yield and grain quality in powdery mildew *Erysiphe graminis* sp. hordei resistant *Hordeum vulgare* cultivar Sultan. Phytopathology 71: 396–399
- Smedegaard-Petersen V and Tolstrup K (1985) The limiting effect of disease resistance on yield. Ann Rev Phytopathol 23: 475–490
- Smith CJ (1996) Accumulation of phytoalexins: Defence mechanism and stimulus response system. New Phytol 132: 1–45
- Somssich IE and Hahlbrock K (1998) Pathogen defence in plants a paradigm of biological complexity. Tr Plant Sci 3: 86–90
- Stadnik MJ and Buchenauer H (1999) Control of wheat diseases by a benzothiadiazole-derivative and modern fungicides. Z Pflanzenk Pflanzen 106: 466–475
- Stout MJ, Brovont RA and Duffey SS (1998a) Effect of nitrogen availability on expression of constitutive and inducible chemical defenses in tomato, *Lycopersicon esculentum*. J Chem Ecol 24: 945–963

- Stout MJ, Fidantsef AL, Duffey SS and Bostock RM (1999) Signal interactions in pathogen and insect attack: Systemic plant-mediated interactions between pathogens and herbivores of the tomato, *Lycopersicon esculentum*. Phys Mol Plant Path 54: 115–130
- Stout MJ, Workman KV, Bostock RM and Duffey SS (1998b) Specificity of induced resistance in the tomato, *Lycopersicon* esculentum. Oecologia 113: 74–81
- Stout MJ, Workman KV, Bostock RM and Duffey SS (1998c) Stimulation and attenuation of induced resistance by elicitors and inhibitors of chemical induction in tomato (*Lycopersicon esculentum*) foliage. Entomol Exp Appl 86: 267–279
- Thaler JS (1999) Jasmonic acid mediated interactions between plants, herbivores, parasitoids, and pathogens: a review of field experiments in tomato. In: Agrawal AA, Tuzun S, and Bent E (eds) Induced plant defenses against pathogens and herbivores: Biochemistry, Ecology, and Agriculture (pp 319–334) The American Phytopathological Society Press, St. Paul, Minnesota
- Thaler JS, Fidantsef AL, Duffey SS and Bostock RM (1997)
  Trade-offs in plant defense against pathogens and herbivores: a field demonstration of chemical elicitors of induced resistance.

  J Chem Ecol 25: 1597–1609
- Tollrian R and Harvell CD (1999) The ecology and evolution of inducible defenses. Princeton University Press, Princeton, New Jersey, USA
- van Loon LC (1985) Pathogenesis-related proteins. Plant Mol Biol 4: 111–116
- van Loon LC (1997) Induced resistance in plants and the role of pathogenesis-related proteins. Eur J Plant Pathol 103: 753–756
- van Loon LC, Bakker PAHM and Pieterse CMJ (1998) Systemic resistance induced by rhizosphere bacteria. Ann Rev Phytopathol 36: 453–483
- van Loon LC and van Strien EA (1999) The families of pathogenesis-related proteins, their activities, and comparative analysis of PR-1 type proteins. Phys Mol Plant Path 55: 85–97
- Van Wees SCM, Pieterse CMJ, Trijssenaar A, Van't Westende YAM, Hartog F and Van Loon LC (1997) Differential induction of systemic resistance in *Arabidopsis* by biocontrol bacteria. Mol Plant-Microbe In 10: 716–724
- Vierheilig H, Alt M, Lange J, Gutrellla M, Wiemken A and Boller T (1995) Colonization of transgenic tobacco constitutively expressing pathogenesis-related proteins by the vesicular-arbuscular mycorrhizal fungus Glomus mosseae. Appplied Environ Microb 61: 3031–3034
- Wasternack C, Ortel B, Miersch O, Kramell R, Beale M, Greulich F, Feussner I, Hause B, Krumm T, Boland W and Parthier B (1998) Diversity in octadecanoid-induced gene expression of tomato. J Plant Phys 152: 345–352
- Wasternack C and Parthier B (1997) Jasmonate-signalled plant gene expression. Tr Plant Sci 2: 302–307
- Weidhase RA, Kramell HM, Lehmann J, Liebisch HW, Lerbs W and Parthier B (1987) Methyl jasmonate-induced changes in the polypeptide pattern of senescing barley leaf segments. Plant Sci 51: 177–186
- Xu Y, Chang PFL, Liu D, Narasimhan ML, Raghothama KG, Hasegawa PM and Bressan RA (1994) Plant defense genes are synergistically induced by ethylene and methyl jasmonate. Plant Cell 6: 1077–1085